

## POSTPARTURIENT HYPOCALCEMIA OF DAIRY COWS: A MODEL FOR THE STUDY OF THE INTERDEPENDENCE OF CA, P<sub>i</sub>, AND MG HOMEOSTASIS

J.-L. Riond<sup>1</sup>, N. Kocabagli<sup>1</sup>, U.E. Spichiger<sup>2</sup> and M. Wanner<sup>1</sup>

<sup>1</sup>Institute of Veterinary Physiology, Division of Animal Nutrition, University of Zurich and  
<sup>2</sup>Centre of Chemical Sensors, Biosensors and (Bio-) Analytical Chemistry, Department of Pharmacy,  
Swiss Federal Institute of Technology, Zurich, Switzerland

### ABSTRACT

Disorders of calcium, phosphorus and magnesium homeostasis in ruminants provide natural models for the study of the physiology and pathophysiology of these minerals. The knowledge that can be acquired with a better understanding of the pathogenesis of these diseases could give useful clues in the puzzle of human osteoporosis. In the present study, the case of parturient paresis of dairy cows is reexamined with a newly developed technique for the measurements of serum ionized magnesium concentrations (Mg<sup>2+</sup>). The concentrations of total magnesium (Mg<sub>tot</sub>), ionized calcium (Ca<sup>2+</sup>), total calcium (Ca<sub>tot</sub>), and inorganic phosphate (P<sub>i</sub>) were also determined in the sera of seventeen 3- to 16-year-old Brown Swiss and crossed Simmental/Red Holstein cows during the periparturient period. In each animal, a transient increase of Mg<sup>2+</sup> and Mg<sub>tot</sub> serum concentrations was observed in association with the transient decrease after parturition of Ca<sup>2+</sup>, Ca<sub>tot</sub> and P<sub>i</sub> serum concentrations. On average, throughout the study, serum Mg<sup>2+</sup> concentrations were 68.5% of those of Mg<sub>tot</sub> whereas serum Ca<sup>2+</sup> concentrations were 52% of those of Ca<sub>tot</sub>. The possible mechanisms involved in the transient increase of Mg<sup>2+</sup> and Mg<sub>tot</sub> serum concentrations are discussed and the relevance of this data for osteoporosis is outlined.

### INTRODUCTION

Among food animal species, sheep and goats are the most adequate candidates for osteoporosis models due to their relative small size. Disorders of calcium, phosphorus and magnesium homeostasis in ruminants of larger size however provide natural models for the study of the physiology and pathophysiology of these minerals (3). Examples in dairy cows are parturient paresis (hypocalcemia), lactation tetany (hypomagnesemia) and postparturient hemoglobinuria (low dietary phosphate) which are diseases whose pathogenesis is influenced or caused by an unbalanced diet. In the case of parturient paresis, the integrated hormonal response has already been extensively studied (20,28,30,35,38). The hypocalcemia which occurs most commonly within the first 48 hours after parturition is related to the sudden drainage of calcium into milk at the onset of lactation. At that time, the cow's body is unable to mobilize enough calcium and the animals become paretic.

Magnesium deficiency causes osteoporosis (12,36,37). It has been postulated that the contribution of magnesium to bone stability occurs by activation of the magnesium-dependent adenosine triphosphatase H<sup>+</sup>-K<sup>+</sup> pump in osteocytes and so by maintaining the pH of the bone extracellular fluid at a slightly higher value (13). Furthermore, in magnesium depletion the secretion of parathyroid hormone (PTH) is impaired and the serum concentrations of 1,25-vitamin D are low (1). Because magnesium concentration is marginally lower in serum and lower in erythrocytes and bone of human patients with postmenopausal or senile osteoporosis than in healthy controls, magnesium may be a contributing factor

*Address for correspondence and reprints:* Dr. J.-L. Riond, Institute of Veterinary Physiology, Division of Animal Nutrition, University of Zurich, Winterthurerstr 260, CH-8057 Zurich, Switzerland.

in the pathogenesis of osteoporosis (8,9,11,29). Magnesium malabsorption may be involved in this process. It has been suggested that supplementation with magnesium should be considered as an adjunct for the prevention and treatment of osteoporosis (39). An exact understanding of the homeostasis of magnesium is thus necessary.

In bovine plasma, about 70% of the magnesium and 50% of the calcium are in the physiologically active free form ( $Mg^{2+}$  and  $Ca^{2+}$ , respectively; 24,41). Approximately 43% of serum calcium is bound to proteins and approximately 10% is complexed with bicarbonate, phosphate, lactate and citrate (14). Similarly, serum magnesium is bound to proteins or organic and inorganic anions (40). Recently, ion-selective electrodes have been designed for determining serum  $Mg^{2+}$  concentration (40,41). For this report, the concentrations of  $Mg^{2+}$ ,  $Ca^{2+}$ , total magnesium ( $Mg_{tot}$ ), total calcium ( $Ca_{tot}$ ), and inorganic phosphorus ( $P_i$ ) were measured in serum during the periparturient period of nonparetic dairy cows.

### ANIMALS, MATERIALS AND METHODS

Seventeen 3- to 16-year-old Brown Swiss and crossed Simmental/Holstein cows were studied. During the sampling period, the cows were housed indoors and fed silage, hay and a conventional concentrate supplement. Two months before parturition, they received poor quality silage and hay in order to reduce energy and calcium intake. The magnesium content of the food was measured and found to adequately cover the daily requirements. Water was provided *ad libitum*. The cows were only partially milked during the 3 days that followed parturition in order to minimize calcium drainage into milk. None of the animals exhibited signs of parturient paresis. Blood samples were obtained from one jugular vein via evacuated tubes. The first samples were taken on one occasion between 15 and 5 days before the expected calving date and once each morning during 6 days after parturition. The blood was allowed to clot, was centrifuged at 1580 g for 10 minutes at 4°C and the serum was aspirated. The serum concentrations of  $Mg^{2+}$  and  $Ca^{2+}$  were quantified within 72 hours. During this period, the samples were maintained at 4°C and both electrolytes were stable. The remaining serum was stored at -20°C for the other analyses. The serum concentrations  $Mg_{tot}$ ,  $Ca_{tot}$ ,  $P_i$ , total protein and albumin were determined by colorimetry with an automated analyser (Cobas Mira, F. Hoffmann-La Roche, Basle, Switzerland), using commercial kits. The analyses were based on the following methods: methylthymol blue for  $Ca_{tot}$ , phosphomolybdate without precipitation of proteins for  $P_i$  and the Biuret reaction for total protein (F. Hoffmann-La Roche), calmagite for  $Mg_{tot}$  (Biomérieux, Lyon, France) and bromocresol green for albumin (Boehringer Mannheim, Germany). Serum potassium (K) concentration was quantitated by flame photometry (Instrumentation Laboratory, Milan, Italy). Serum osmolarity was determined by depression of the freezing point (Knauer automatic semi-micro osmometer, Berlin, Germany). Serum  $Ca^{2+}$  and  $Mg^{2+}$  concentrations were quantitated using ion selective electrodes (AVL 988-4 prototype apparatus, AVL List GmbH, Graz, Austria) with internal quality and accuracy controls and were corrected for the mean physiological pH 7.4 (37°C) by linear regression. Changes with time in the parameters  $Mg^{2+}$ ,  $Mg_{tot}$ ,  $Ca^{2+}$ ,  $Ca_{tot}$ ,  $P_i$ , total protein, albumin, K and osmolarity were examined by repeated measurement analysis of variance using the version 6.08 of the main frame computer-implemented SAS procedure ANOVA (SAS Institute Inc, Cary, NC, USA). For each parameter, the values for the days postpartum were contrasted with those of the prepartum period or between adjacent days by use of the options contrast and profile, respectively. The level of significance was fixed at 0.05.

### RESULTS

In each animal, a transient increase of serum  $Mg^{2+}$  and  $Mg_{tot}$  concentrations was observed in association with the transient decrease after parturition of serum  $Ca^{2+}$ ,  $Ca_{tot}$ , and  $P_i$  concentrations. The situation is illustrated for one 8-year-old cow (Fig 1). Plots for individual animals indicated that the concentration profiles of serum  $Mg^{2+}$  and  $Mg_{tot}$  and of  $Ca^{2+}$  and  $Ca_{tot}$  were parallel and that the duration of the increase of  $Mg^{2+}$  and  $Mg_{tot}$  was related to the duration of the decrease of  $Ca^{2+}$  and  $Ca_{tot}$ . For both serum magnesium and calcium, the degree of complexation remained constant during the hypocalcaemic

episode. On average, serum  $Mg^{2+}$  concentrations were 68.5% of those of  $Mg_{tot}$  whereas serum  $Ca^{2+}$  concentrations were 52.0% of those of  $Ca_{tot}$ . In the serum of almost all animals, the  $P_i$  concentration profiles were parallel to those of  $Ca^{2+}$  and  $Ca_{tot}$ . When compared with reference laboratory values, the serum concentrations of  $Ca^{2+}$ ,  $Ca_{tot}$  and  $P_i$  of some cows dropped to values below the normal range whereas the peak increase for  $Mg^{2+}$  and  $Mg_{tot}$  always stayed within the physiological norm. When all cows were considered, the repeated measurement analysis of variance indicated that the serum concentrations of  $Ca^{2+}$ ,  $Ca_{tot}$ , and  $P_i$  were significantly decreased during day 1 and day 2 postpartum, whereas those of  $Mg^{2+}$  and  $Mg_{tot}$  were significantly increased. On day 3, serum  $Ca_{tot}$  concentrations were still significantly decreased. Serum osmolarity and albumin, total protein and K serum concentrations remained unchanged throughout the course of the study.

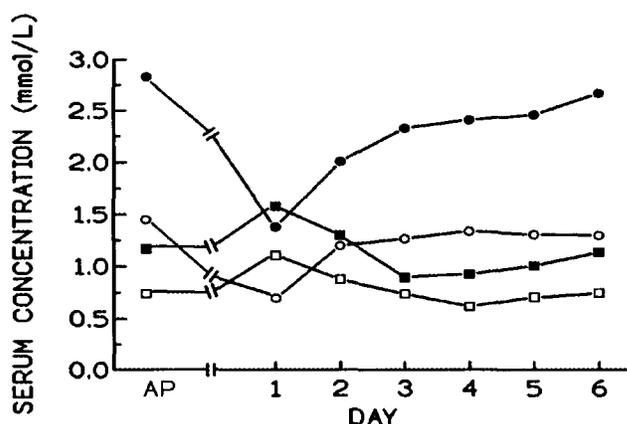


FIG. 1 Serum concentrations of  $Ca_{tot}$  (●),  $Ca^{2+}$  (○),  $Mg_{tot}$  (■) and  $Mg^{2+}$  (□) in one 8-year-old Brown Swiss cow during the periparturient period.

## DISCUSSION

The parallel profiles of the serum concentrations of  $Mg^{2+}$  and  $Mg_{tot}$  during the course of the mild nonclinical hypocalcemia indicate that the free concentration may be predicted from the total concentration. In mild hypocalcemia, the concentrations of the ligands of serum magnesium are not altered and the ratio  $Mg^{2+}/Mg_{tot}$  was thus maintained. Similarly, the parallelism of the profiles of the serum concentrations of  $Ca^{2+}$  and  $Ca_{tot}$  of this report confirmed the findings of other studies which have included paretic cows (4,23,24).

Serum osmolarity and serum concentrations of albumin and total protein remained unchanged during the hypocalcemic episode indicating that no dehydration occurred. During the periparturient period, water consumption has never been measured but appetite is reduced (35,38). Dehydration caused by decreased water intake was suspected because water intake is associated with food intake in healthy pigmy goats (32). Hemoconcentration has been observed in animals with clinical signs of parturient paresis (34). Because no dehydration was present in the cows of this study, an adjustment of  $Ca_{tot}$  concentration for albumin concentration was not necessary.

The milk/serum concentration ratios are 10 to 15 for  $Ca_{tot}$ , 5 for  $Mg_{tot}$  and 20 for  $P_i$  (38). The transiently increased serum concentrations of  $Mg^{2+}$  and  $Mg_{tot}$  associated with the decreased  $Ca^{2+}$  and  $Ca_{tot}$  is remarkable in light of magnesium drainage into milk. This phenomenon has been observed in several studies related with postparturient paresis (4,18,44) and during short term fasting which is characterized by decreased serum  $Ca_{tot}$  concentration (17,18). Conversely, a decreased  $Mg_{tot}$  was reported after administration of  $Ca_{tot}$ -elevating doses of vitamin D metabolites (43,44). This effect may presumably be attributable to the hypercalcemia that is induced (30). These findings strongly suggest

that one or some of the mechanisms involved in the regulation of calcium and  $P_i$  homeostasis in cows also affect the serum concentrations of  $Mg^{2+}$  and  $Mg_{tot}$ . In support of this hypothesis, urinary magnesium is transiently decreased during the hypocalcemic period that follows parturition in dairy cows, suggesting increased tubular reabsorption (18). Furthermore, in the kidney of rats, the segments 1-34 of bovine PTH and synthetic PTH-related protein (PTHrP) infused via osmotic minipumps stimulate the reabsorption of magnesium (31). Also, infusion of bovine PTH for 60 minutes promotes the reabsorption of magnesium in the kidney of thyroparathyroidectomized hamsters (7). Finally, infusion of PTH in dairy cows during the last trimester of pregnancy induces an increase of serum  $Mg_{tot}$  concentration (15). Thus, the effect on the kidney of increased serum PTH induced by the hypocalcemia could be one of the factors involved in the transient increase of serum  $Mg^{2+}$  and  $Mg_{tot}$  concentrations observed in periparturient cows.

Assuming a milk magnesium concentration of 4.5 mmol/L and a milk production of 10 L, approximately 1 g of magnesium is excreted into milk. Using the data from Halse (18), it may be calculated that the magnesium loss in milk is compensated with the 1 g of magnesium reabsorbed in the kidney. However, the increase of serum  $Mg_{tot}$  concentration of 0.20 mmol/L corresponds to a magnesium increase of 1 g in the extracellular fluid volume. Magnesium must thus be mobilized from another source which may be bone, the gastro-intestinal tract, a shift of magnesium from the intracellular compartment to the extracellular compartment or a combination of the 3 factors.

As 60% of the body magnesium is stored in bone, mobilisation from bone at the time of parturition could contribute to the observed increase of  $Mg^{2+}$  and  $Mg_{tot}$  (27). Bone magnesium is present in the bone extracellular fluid, in the hydration shell of the hydroxyapatite crystals and on the crystal surfaces, but not in the crystal interior (16). PTH administration induces in a first stage and without the activation of osteoclasts the release of bone calcium from a rapidly turning over pool, presumably the bone extracellular fluid and the hydration shell on the crystal surfaces (22), and could thus also induce magnesium release. It is difficult to evaluate how much of the 150 to 200 g of magnesium of the cows' bones may be mobilized by this mechanism. In a later phase (within 24 hours), after stimulation of PTH release or PTH administration which activates osteoclasts and results in increased bone resorption (5), more magnesium may be released from bone. However, because the Ca:Mg ratio is about 50:1, a large amount of bone has to be remodeled to yield a significant amount of magnesium.

Vitamin D and its hydroxylated metabolites enhance intestinal magnesium absorption in rats and in humans (18). In the ovine and bovine rumen, an active transport has been demonstrated for the absorption of magnesium (25,26). Whether Vitamin D and its metabolites are involved in the absorption of magnesium in ruminants is unknown. As vitamin D concentration increases during postparturient hypocalcemia (30,38), it could possibly be involved in an enhancement of magnesium absorption and thus contribute to the observed transient increase of  $Mg^{2+}$  and  $Mg_{tot}$ .

A shift of magnesium from the intracellular compartment to the extracellular compartment during the course of hypocalcemia may also be involved in the observed transient increase of  $Mg^{2+}$  and  $Mg_{tot}$ . Because serum K concentration remained constant, the involuting uterus was very probably not the source of the elevation of serum  $Mg^{2+}$  and  $Mg_{tot}$ . Magnesium is only second to K as the main intracellular ion (33). A reduced affinity of magnesium for plasma proteins during hypocalcemia could also be involved.

At the onset of lactation, the portion of the large amount of the PTH-related protein produced in the mammary gland that is released in the blood stream may also play a role in magnesium homeostasis, because the biological activity of PTH-related protein is identical to that of PTH (2,21). Other hormones including some of the hormones involved in stress affect magnesium homeostasis: calcitonin, growth hormone, aldosterone, antidiuretic hormone, glucagon,  $\beta$ -adrenergic agonists and insulin (6,10). However, magnesium is less rigidly regulated than calcium and a specific magnesium-regulating hormone has not been identified (42).

In conclusion, the observations on magnesium concentration during the course of hypocalcemia of parturient paresis of dairy cows raise interesting questions. A more complete understanding of the pathogenesis of this disease and other metabolic diseases of ruminants may provide clues on the homeostatic regulation of this mineral. This in turn could shed some light into some aspects of bone

physiology and pathophysiology and consequently allow to design better strategies for the treatment and the prevention of osteoporosis.

## REFERENCES

1. Abbott, L., Nadler, J., Rude, R.K. Magnesium deficiency in alcoholism: Possible contribution to osteoporosis and cardiovascular disease in alcoholics. *Alcohol Clin Exp Res* 18: 1076-1082; 1994.
2. Barlet, J.-P., Abbas, S.K., Care, A.D., Davicco, M.-J., Rouffet, J. Parathyroid hormone-related peptide and milking-induced phosphaturia in dairy cows. *Acta Endocrinol* 129: 332-336; 1993.
3. Blood, D.C., Radostits, O.M. Metabolic diseases. *Veterinary Medicine: a textbook of the diseases of cattle, sheep, pigs, goats and horses*. 7th ed. London: Baillière Tindall: 1989; 1100-1149.
4. Blum, J.W., Ramberg, C.F., Johnson, K.G., Kronfeld, D.S. Calcium, magnesium, phosphorus, and glucose in plasma from parturient cows. *Am J Vet Res* 33: 51-56; 1972.
5. Braak, A.E., van de Klooster, A.T. van't, Hal-van Gestel, J.C. van, Malestein, A. Influence of stage of lactation and calcium level of the ration on mobilisation rate of calcium and excretion of hydroxyproline in urine in dairy cows. Studies with Na<sub>2</sub>EDTA infusions in monozygotic twins. *Zbl Vet A - J Vet Med A* 31: 725-739; 1984.
6. Brink, E.J., Beynen, A.C. Nutrition and magnesium absorption: a review. *Prog Food Nutr Sci* 16: 125-162; 1992.
7. Burnatowska, M.A., Harris, C.A., Sutton, R.A.L., Dirks, J.H. Effects of PTH and cAMP on renal handling of calcium, magnesium, and phosphate in the hamster. *Am J Physiol* 233: F514-F518; 1977.
8. Cohen, L. Recent data on magnesium and osteoporosis. *Magnesium Res* 1: 85-87; 1988.
9. Cohen, L., Kitzes, R. Infrared spectroscopy and magnesium content of bone mineral in osteoporotic women. *Isr J Med Sci* 17: 1123-1125; 1981.
10. De Rouffignac, C., Mandon, B., Wittner, M., Di Stefano, A. Hormonal control of renal magnesium handling. *Min Electrolyte Metab* 19: 226-231; 1993.
11. Driessens, F.C.M., Steidl, L., Ditmar, R. Magnesium, calcium and zinc status in different forms of osteoporosis. *Magnesium Bull* 12: 158-160; 1990.
12. Driessens, F.C.M., Verbeeck, R.M.H., Dijk, J.W.E., van, Borggreven, J.M.P.M. Response of plasma calcium and phosphate to magnesium depletion. A review and its physiological interpretation. *Magnesium Bull* 9: 193-201; 1987.
13. Driessens, F.C.M., Verbeeck, R.M.H., Dijk, J.W.E., van. A systemic approach to the oral problem of mandibular resorption. *Bull Group Int Rech Sci Stomat Odontol* 32: 127-137; 1989.
14. Forman, D.T., Lorenzo, L. Ionized calcium: Its significance and clinical usefulness. *An Clin Lab Sci* 21: 297-304; 1991.
15. Goff, J.P., Littledike, E.T., Horst, R.L. Effect of synthetic bovine parathyroid hormone in dairy cows: Prevention of hypocalcemic parturient paresis. *J Dairy Sci* 69: 2278-2289; 1986.
16. Green, J. The physicochemical structure of bone: Cellular and noncellular elements. *Min Electrolyte Metab* 20: 7-15; 1994.
17. Halse, K. Changes in serum calcium and magnesium in cows subjected to short periods of fasting. *Proc VIII International Grassland Congress, Reading, Berkshire, England*. Oxford, UK: The Alden Press: 1961; 553-558.
18. Halse, K. Calcium effects on renal conservation of magnesium in cows. *Acta Vet Scand* 25: 213-228; 1984.
19. Hardwick, L.L., Jones, M.R., Brautbar, N., Lee, D.B.N. Magnesium absorption: Mechanisms and the influence of vitamin D, calcium and phosphate. *J Nutr* 126: 13-23; 1991.
20. Horst, R.L., Goff, J.P., Reinhardt, T.A. Calcium and vitamin D metabolism in the dairy cow. *J Dairy Sci* 77: 1936-1951; 1994.
21. Kocabagli, N., Riond, J.-L., Spichiger, U.E., Wanner, M. Parathyroid hormone-related protein and calcium homeostasis during the periparturient period of dairy cows. *Am J Vet Res* 56: 380-385; 1995.
22. Kronenberg, H.M. Parathyroid Hormone: Mechanism of Action. Favus, M.J., ed. *Primer on the Metabolic Bone Diseases and Disorders of Mineral Metabolism*. New York: Raven Press: 1993; 58-60.

23. Kwart, C., Björnsell, K.A., Larsson L. Parturient paresis in the cow: serum ionized calcium concentrations before and after treatment with different calcium solutions - Classification of different degrees of hypo- and hypercalcemia. *Acta Vet Scand* 23: 184-196; 1982.
24. Lincoln, S.D., Lane, V.M. Serum ionized calcium concentration in clinically normal dairy cattle, and changes associated with calcium abnormalities. *J Am Vet Med Assoc* 197: 1471-1474; 1990.
25. Martens, H. Saturation kinetics of magnesium efflux across the rumen wall in heifers. *Br J Nutr* 49: 153-158; 1983.
26. Martens, H., Harmeyer, J., Michael, H. Magnesium transport by isolated rumen epithelium of sheep. *Res Vet Sci* 24: 161-168; 1978.
27. Matsui, T., Yano, H., Kawabata, T., Harumato, T. The effect of suppressing bone resorption on Mg metabolism in sheep (*Ovis aries*). *Comp Biochem Physiol (A)* 107: 233-236; 1994.
28. Potts, J.T., Buckle, R.M., Sherwood, L.M., Ramberg, C.F., Mayer, C.P., Kronfeld, D.S., Deftos, L.J., Care, A.D., Aurbach, G.D. Control of secretion of parathyroid hormone. Talmage, R.V., Bélanger L.F., Clark I., eds. *Parathyroid Hormone and Thyrocalcitonin (Calcitonin)*. Proc Third Parathyroid Conference, Montreal. Amsterdam: Excerpta Medica: 1968; 407-416.
29. Reginster, J.Y., Strause, L., Deroisy, R., Lecart, M.P., Saltman, P., Franchimont, P. Preliminary report of decreased serum magnesium in postmenopausal osteoporosis. *Magnesium* 8: 106-109; 1989.
30. Reinhardt, T.A., Horst, R.L., Goff, J.P. Calcium, phosphorus, and magnesium homeostasis in ruminants. *Vet Clin N Am-Food Anim Pr* 4: 331-350; 1988.
31. Rizzoli, R., Caverzasio, J., Chapuy, M.C., Martin, T.J. Bonjour, J.P. Role of bone and kidney in parathyroid hormone-related peptide-induced hypercalcemia in rats. *J Bone Min Res* 4: 759-765; 1989.
32. Rossi, R., Scharrer, E. Circadian patterns of drinking and eating in pigmy goats. *Physiol Behav* 51: 895-897; 1992.
33. Ryan, M.P. Interrelationships of magnesium and potassium homeostasis. *Min Elektrolyte Metab* 19: 290-295; 1993.
34. Rydberg, C. Hämatologische und blutchemische Untersuchungen von klinisch gesunden sowie an primärer Ketose und perperaler Parese erkrankten Rindern *Acta Vet Scand Suppl* 27: 41-72; 1969.
35. Sechen, S.J., Bremel, R.D., Jorgensen, N.A. Prolactin, estradiol, and progesterone changes in paretic and nonparetic cows during the periparturient period. *Am J Vet Res* 49: 411-416; 1988.
36. Seelig, M.S. Abnormal bone in magnesium deficiency. Magnesium deficiency in the pathogenesis of disease. New York: Plenum Medical Book Company: 1980; 285-329.
37. Seelig, M.S. Interrelationship of magnesium and estrogen in cardiovascular and bone disorders, eclampsia, migraine and premenstrual syndrome. *J Am Coll Nutr* 12: 442-458; 1993.
38. Shappell, N.W., Herbein, J.H., Deftos, L.J., Aiello, R.J. Effects of dietary calcium and age on parathyroid hormone, calcitonin and serum and milk minerals in the periparturient dairy cow. *J Nutr* 117: 201-207; 1987.
39. Sojka, J.E., Weaver, C.M. Magnesium supplementation and osteoporosis. *Nutr Rev* 53: 71-74; 1995.
40. Spichiger, U.E., Eugster, R., Citterio, D., Li, H., Schmid, A., Simon, W. Magnesium activity measurements: facts and enthusiasm. Golf, S., Dralle, D., Vecchiet, L., eds. *Magnesium 1993*, London: John Libbey & Company Ltd: 1994; 49-60.
41. Spichiger, U.E., Wild, R. Magnesium the forgotten ion. Porta J et al., eds. *Proc Int Mg<sup>2+</sup>-days*, Bad Radkersburg, Austria. Graz, Austria: Leykam Buchgesellschaft: 1994; 19-37.
42. Toffaletti, J., Cooper, D.L., Lobaugh, B. The response of parathyroid hormone to specific changes in either ionized calcium, ionized magnesium, or protein-bound calcium in humans. *Metabolism* 40: 814-818; 1991.
43. Zepperitz, H., Gürtler, H. Ionisiertes Calcium und Gesamtcalcium im Blut von Rindern, Schafen, Schweinen und Pferden verschiedener Alters- und Reproduktionsstadien und Nutzungsrichtungen. *Berl Mün Tierarztl Wochenschr* 105: 328-332; 1992.
44. Zepperitz, H., Gürtler, H., Schäfer, M., Glatzel, E. Einfluss einer Prophylaxe der Gebärparese mit 1-alpha-Hydroxycholecalciferol auf die Konzentrationen an ionisiertem Calcium im Blut und weiteren Mineralstoffen im Blutplasma bei der Milchkuh. *Monatsh Veterinärmed* 49: 13-21; 1994.